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Optimum Temperatures for the Peak Growth of Some Selected Bacterial Fish Pathogens

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Abstract

The optimal growth temperatures for 24 strains of 15 bacterial fish pathogens were determined under uniform cultural conditions. For *Flavobacterium psychrophila*, peak growth temperature (t_{peak}) was $21 \pm 0.4^\circ\text{C}$ for the strain tested. *Flavobacterium columnaris* exhibited 29.8 ± 0.2 and $29.9 \pm 0.3^\circ\text{C}$, respectively, as t_{peaks} for two strains. The t_{peaks} of *Flavobacterium branchiophilum* were 24.9 ± 0.2 and $24.1 \pm 0.5^\circ\text{C}$, respectively, for two strains. Three strains of *Edwardsiella tarda* were examined and they showed t_{peaks} at 36.1 ± 0.4 , 35.5 ± 0.2 and $36 \pm 0.3^\circ\text{C}$, respectively. *Citrobacter freundii* showed its t_{peak} at $36.1 \pm 0.4^\circ\text{C}$. For *Aeromonas hydrophila*, two strains were tested and it was found that t_{peaks} were 32.9 ± 0.6 and $34.8 \pm 0.5^\circ\text{C}$, respectively. The t_{peaks} of *Aeromonas salmonicida* were 26.1 ± 0.4 and $25.6 \pm 0.2^\circ\text{C}$, respectively. Temperature $27.8 \pm 0.3^\circ\text{C}$ was found as t_{peak} of a *Vibrio anguillarum* strain while *Vibrio ordalii* strain exhibited it at $26.5 \pm 0.4^\circ\text{C}$. Peak growth at temperatures of 26.7 ± 0.5 and $27.6 \pm 0.4^\circ\text{C}$, respectively, was found for *Photobacterium damsela* (formerly *Pasteurella piscicida*). In *Pseudomonas fluorescens*, the peak growth occurred at temperatures of 29.9 ± 0.2 and $28.5 \pm 0.3^\circ\text{C}$, respectively. *Pseudomonas* sp. showed t_{peaks} at 24.9 ± 0.1 and $29.2 \pm 0.5^\circ\text{C}$, respectively. The peak growth for the two strains of *Lactococcus garvieae* (formerly *Enterococcus seriolicida*) was at temperatures of 29.7 ± 0.4 and $31.1 \pm 0.2^\circ\text{C}$, respectively where the *Streptococcus* sp. showed t_{peak} at $28.3 \pm 0.5^\circ\text{C}$.

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Introduction

Bacterial fish disease is one of the major constraints in the development of aquaculture. Particularly in intensive aquaculture, basic hygiene which is necessary for maintaining good water quality is often overlooked and may lead to the start of a disease cycle (McCarthy 1977). Water temperature is one of the vital factors for growth, reproduction, disease and other biological activities of fish. The progress of fish disease is influenced by environmental temperature. *Flavobacterium psychrophila* has been well documented as a fish pathogen (Borg 1960). *Flavobacterium columnaris* is the causative agent of columnaris disease (Davis 1922). The effect of water temperature on columnaris disease in Oriental weather fish was studied by Wakabayashi & Egusa (1972). All of the exposed fish held at 20-35°C died. The relationship of water temperature with the number of fish infections induced by *Aeromonas hydrophila* had been reported (Groberg et al. 1978). The seasonality in the occurrence of disease is thought to be due to the change in environmental temperature. In disease, environmental temperature can influence not only fish but also the pathogen. Many bacteria are known to cause fish disease and their growth temperatures are different from genus to genus. The most appropriate temperature for the growth of pathogen is not the same with the temperature occurring disease and pathogenicity. It is known that the environmental temperature which causes disease has a tendency to be lower than the optimal growth temperature of pathogen but the details are not known. In this study we compared the optimal growth temperature of some important bacterial fish pathogens. Knowledge of the factors affecting growth (temperature, NaCl concentration for marine bacterium, pH etc.) is indispensable before embarking on susceptibility testing. For the production of highly effective bacterins, it is essential to know the cultural conditions of a particular bacterium.

However, many investigators worked on different bacterial fish pathogens and in most cases only a single bacterium was completely examined. Use of different media, different growth conditions and a variety of assay made the literature reports very difficult to interpret and compare. In this study, optimal growth temperatures of some selected strains of bacterial fish pathogens were determined under the uniform cultural conditions.

Materials and Methods

Bacterial strains

A total of 24 strains, one to three from each of 15 different bacterial species of fish or pond water origin, were selected for this study. The details of the bacterial strains studied are summarized in [table 1](#). All these bacterial strains were collected from France, Japan, KSA and USA. After collection, they were preserved at -80°C on a long term basis. Stock cultures of the organism were maintained on tryptone-soya broth “Nissui” (TSB) or tryptone yeast extract (TYE) broth at 18 or 25°C incubation temperature. The TYE broth was used for the culture of *F. psychrophila*, *F. columnaris* and *F. branchiophilum*. An extra 1.5% NaCl was added to TSB for *V. anguillarum*, *V. ordalii*, *P. damsela* and *L. garvieae* while the rest of the bacteria were cultured in TSB. For inoculum, a loopful of actively growing broth culture was inoculated in 18 ml of each broth in an L-shaped tube and subsequently incubated at different points of temperature gradient incubator (TGI: Toyo Co.,Tn-3), over a range of 3-48°C. The shaking speed was 30•min⁻¹. Media composition: TSB = peptone (17 g), bean peptone (3.0 g), NaCl (5.0 g), glucose (2.5 g), KHPO₄ (2.5 g) in a total amount of 30 g media dissolved in 1000 ml distilled water having pH 7.3±0.1. TYE = tryptone (0.4%), yeast extract (0.05%), CaCl₂•2H₂O (0.02%), MgSO₄•7H₂O (0.05%) and pH 7.2±0.1.

Determination of growth

Growth of organisms was monitored by measuring the optical density of the culture by a spectrophotometer (Hitachi, U-2000) at 480 nm. In a pre-study, time course effect on the growth of several bacteria at different temperatures was determined and it was found that similar growth patterns were followed up to an OD 0.8 within log phase culture ([Tables 2 and 3](#)). So, samples were collected when the OD was lower than 0.6 (within log phase) to determine the growth after respective incubation hours of different strains.

Results

The main objective peak growth optimum temperatures of 24 strains of 15 bacterial species were determined from the mean of three

Table 1. Optimum growth temperatures of different bacterial fish pathogen strains used in this study

Bacterial species	Strain/ Isolation year	Source (Fish / Pond water)	Locality	Initial culture temperature (°C)	^a Peak growth temperature (°C)	Growth range temperature (°C)	Incubation period (h)
<i>Flavobacterium psychrophila</i>	FPC805/1988	<i>Oncorhynchus mykiss</i>	France	18	21.0±0.4	12-24	20
<i>Flavobacterium columnaris</i>	FPC74/66	<i>Misgurnus anguillicaudatus</i>	Japan	25	29.8±0.2	22-23	22
	FPC667/87	<i>Carassius auratus</i>	Japan	25	29.9±0.3	19-33	18
<i>Flavobacterium branchiophilum</i>	ATCC35035/77	<i>Oncorhynchus masou</i>	Japan	18	24.9±0.2	13-30	40
	ATCC35036/78	<i>Oncorhynchus tshawytscha</i>	USA	18	24.1±0.5	12-28	53
<i>Edwardsiella tarda</i>	FPC22/80	<i>Anguilla japonica</i>	Japan	25	36.1±0.4	24-41	6
	FPC611/81	Eel-pond water	Japan	25	35.5±0.2	23-40	6
	FPC802/79	<i>Oreochromis niloticus</i>	Japan	25	36.0±0.3	26-40	6
<i>Citrobacter freundii</i>	FPC349/ -	Eel-pond water	Japan	25	36.1±0.4	27-41	5
<i>Aeromonas hydrophila</i>	FPC868/78	<i>Anguilla japonica</i>	Japan	25	32.9±0.6	20-39	6
	AHC02/2001	Hybrid tilapia	KSA	25	34.8±0.5	19-42	6
<i>Aeromonas salmonicida</i>	NCMB2020/ -	-	-	18	26.1±0.4	17-33	13
	FPC367/78	<i>Oncorhynchus mykiss</i>	Japan	18	25.6±0.2	18-31	14
<i>Vibrio anguillarum</i>	FPC92/66	<i>Plecoglossus altivelis</i>	Japan	25	27.8±0.3	19-35	11
<i>Vibrio ordalii</i>	FPC676/74	<i>Plecoglossus altivelis</i>	Japan	25	26.5±0.4	19-35	9
<i>Photobacterium damsela</i>	FPC851/87	<i>Seriola quinqueradiata</i>	Japan	25	26.7±0.5	23-32	35

Table 1. Optimum growth temperatures of different bacterial fish pathogen strains used in this study (continued)

Bacterial species	Strain/ Isolation year	Source (Fish / Pond water)	Locality	Initial culture temperature (°C)	^a Peak growth temperature (°C)	Growth range temperature (°C)	Incubation period (h)
<i>Photobacterium damsela</i>	FPC852/87	<i>Seriola quinqueradiata</i>	Japan	25	27.6±0.4	22-31	22
<i>Pseudomonas fluorescens</i>	FPC96/73	<i>Cyprinus carpio</i>	Japan	25	29.9±0.2	19-34	12
<i>Pseudomonas</i> sp. (fluorescens-like)	FPC348/-	<i>Salvelinus pluvius</i>	Japan	25	28.5±0.3	18-32	13
	FPC78/73	<i>Cyprinus carpio</i>	Japan	25	24.9±0.1	15-29	11
<i>Pseudomonas</i> sp. (putida-like)	FPC951/94	<i>Plecoglossus altivelis</i>	Japan	25	29.2±0.5	23-31	7
<i>Lactococcus garvieae</i>	FPC137/75	<i>Seriola quinqueradiata</i>	Japan	25	29.7±0.4	16-42	9
	FPC871/90	<i>Paralichthys olivaceus</i>	Japan	25	31.1±0.2	20-38	14
<i>Streptococcus</i> sp.	AHC92/92	Hybrid tilapia	KSA	25	28.3±0.5	15-40	14

^a=mean ± standard deviation (SD) of three replicates

Table 2. Time course effect on the growth response of *Flavobacterium psychrophila* at different temperatures

Temperature (°C)	Growth Optical Density					
	11 h	14h	17 h	20 h	23 h	26 h
12.5	0	0	0.02	0.03	0.13	0.21
15.5	0	0	0.07	0.19	0.43	0.75
18.5	0	0.04	0.17	0.40	0.54	0.92
19.5	0	0.08	0.23	0.44	0.72	1.04
20.7	0	0.12	0.28	0.51	0.80	1.05
21.6	0	0.07	0.19	0.38	0.51	0.73

Table 3. Time course effect on the growth response of *Aeromonas hydrophila* (a representative of mesophilic bacteria) at different temperatures

Temperature (°C)	Growth Optical Density					
	0 h	4h	5 h	6 h	7 h	8h
30.3	0	0.09	0.20	0.39	0.55	0.73
31.8	0	0.14	0.28	0.53	0.80	1.10
33.2	0	0.17	0.32	0.58	0.84	1.10
34.6	0	0.13	0.25	0.52	0.73	1.02
35.9	0	0.11	0.27	0.46	0.57	0.82
37.2	0	0.07	0.22	0.37	0.45	0.59

replications in each case. The growth temperature ranges were also recorded though the incubation period was limited. The effect of temperature on the growth of all investigated strains is summarized in [table 1](#). Growth of all the strains was profoundly affected by incubation temperature as a whole. Peak growth temperature (t_{peak}) of *F. psychrophila* strain was $21 \pm 0.4^\circ\text{C}$. The growth temperature range was $12\text{--}24^\circ\text{C}$ for an incubation period of 20 h. The t_{peaks} for the two strains of *F. columnaris* were 29.8 ± 0.2 and $29.9 \pm 0.3^\circ\text{C}$, respectively. Growth temperature ranges were $19\text{--}33^\circ\text{C}$ for 18–22 h culture. The t_{peaks} of *F. branchiophilum* were 24.9 ± 0.2 and $24.1 \pm 0.5^\circ\text{C}$, respectively, where two strains were tested. A 40–53 h culture showed growth temperature ranges as $12\text{--}30^\circ\text{C}$. Three strains of *E. tarda* were examined and they showed t_{peaks} at 36.1 ± 0.4 , 35.5 ± 0.2 and $36 \pm 0.3^\circ\text{C}$, respectively. Temperatures $23\text{--}41^\circ\text{C}$ were found as growth ranges for 6 h cultivation period. A strain of *C. freundii* stood at $36.1 \pm 0.4^\circ\text{C}$ for its t_{peak} . Growth temperature range was $27\text{--}41^\circ\text{C}$ for 5 h. For *A. hydrophila* two strains were tested and it was found that t_{peaks} were 32.9 ± 0.6 and $34.8 \pm 0.5^\circ\text{C}$, respectively. The growth temperature ranges were $19\text{--}42^\circ\text{C}$ for 6 h. The peak growth temperatures for two strains of *A. salmonicida* were at 26.1 ± 0.4 and $25.6 \pm 0.2^\circ\text{C}$, respectively. The growth temperature ranges were $17\text{--}33^\circ\text{C}$ for 13–14 h cultures. The temperature $27.8 \pm 0.3^\circ\text{C}$ was found as t_{peak} of *V. anguillarum* strain while *V. ordalii* strain gave t_{peak} at $26.5 \pm 0.4^\circ\text{C}$. In both cases, growth temperature range was $19\text{--}35^\circ\text{C}$ at 9–11

h incubation period. Peak growth at temperatures of 26.7 ± 0.5 and $27.6 \pm 0.4^\circ\text{C}$, respectively, was found in *P. damsela* for two strains. After 22-35 h cultivation, growth temperature ranges were between 22 and 32°C . In *P. fluorescens* peak growth was evident at temperatures of 29.9 ± 0.2 and $28.5 \pm 0.3^\circ\text{C}$, respectively, for two strains. Temperature ranges for growth were at $18\text{--}34^\circ\text{C}$ after 12-13 h incubation. The strain of *Pseudomonas* sp. (capsulated, fluorescens-like) stood for t_{peak} at $24.9 \pm 0.1^\circ\text{C}$ while the putida-like *Pseudomonas* sp. strain showed it at $29.2 \pm 0.5^\circ\text{C}$. Capsulated one showed the growth temperature range as $15\text{--}29^\circ\text{C}$ for 11 h while the putida-like gave $23\text{--}31^\circ\text{C}$ at 7 h culture. Temperatures of 29.7 ± 0.4 and $31.1 \pm 0.2^\circ\text{C}$, respectively, showed the peak growth for two strains of *L. garvieae*. The growth temperature ranges were $16\text{--}42^\circ\text{C}$ for 9-14 h incubation period. A strain of *Streptococcus* sp. gave peak growth temperature at $28.3 \pm 0.5^\circ\text{C}$ and growth range as $15\text{--}40^\circ\text{C}$ for 14 h of incubation.

Discussion

Temperature is a key factor for controlling the rate of development of microbial populations, a modulation of enzyme synthesis by the growth temperature having been observed in several microorganisms (Gugi et al. 1991). Pacha (1968) noted that the growth optimal of *F. psychrophila* was around 20°C and growth was absent at 30°C . These results fully coincided with our present findings. According to Holt et al. (1989) optimal growth of *F. psychrophila* was at 15°C . Nomura & Ohara (1994) noted that the optimum growth temperature of *F. columnaris* was 25°C . Our results showed a higher optimal growth temperature. The *F. branchiophilum* grew at temperatures between 10 and 25°C , though some strains grew at a temperature of 5 or 30°C (Wakabayashi et al. 1989). We observed a lightly higher temperature affinity for the growth, compared to literature.

Our *E. tarda* results followed the findings of Farmer & McWhorter (1984) where they noted the optimum growth temperature as 37°C . The present *C. freundii* fully satisfied the results in Bergey's manual (Sakazaki 1984). Optimal temperature of *A. hydrophila* is considered to be 28°C but Statner et al. (1988) mentioned a higher temperature than that. Rouf & Rigney (1971) noted it at 35°C . These results are parallel to our findings. Popoff (1984) found optimum growth temperature for *A. salmonicida* at $22\text{--}25^\circ\text{C}$, lower than the present work. Guerin-Faubleee et al. (1995) found the optimum temperature range for *V. anguillarum* at $29.7\text{--}34^\circ\text{C}$. Our

results verily differed with [Larsen \(1984\)](#) where optimum temperature was at 25°C. The *V. ordalii* exhibited a higher growth temperature than the results, 15-22°C found by [Schiewe et al. \(1981\)](#). According to [Hashimoto et al. \(1985\)](#) growth optimum temperature range of *P. damsela* was 22.5-30°C. It is thought that infection takes place in sea water at a temperature of approximately 25°C ([Yasunaga et al. 1983](#)). Our results supported these results. It is speculated that the growth optimal temperature might have some relation to infection. Literature showed that *P. fluorescens* grew well on nutrient agar at 22-25°C. Putida-like *Pseudomonas* sp. causes bacterial haemorrhagic ascites of ayu, *Plecoglossus altivelis* ([Wakabayashi et al. 1996](#)). Capsulated, fluorescence-like *Pseudomonas* sp. was isolated from carp. The peak optimal temperatures in different *Pseudomonas* spp. were varied. [Kusuda et al. \(1991\)](#) stated that *L. garvieae* grew at 10-45°C. The optimum temperature for the growth of B-hemolytic streptococcus was 25-30°C ([Ohnishi & Jo 1986](#)). For *Streptococcus* sp., [Al-Harbi \(1994\)](#) reported the optimum growth temperature at 30°C and in our study it was at 28.3±0.5°C. It is noted that due to incubation time limit, we did not compare the present results of growth temperature ranges with the other findings.

Strains and temperature generally exerted a considerable influence on the growth of bacteria. The impact of temperature on growth of bacteria is in general highly significant. The appropriate temperatures for growth of bacterial fish pathogens differ in their genera and the range was 21±0.4 to 36.1±0.4°C. The optimum temperatures for most of the examined bacteria were more or less similar to the literature available but some of them varied. In some cases, slightly higher temperatures than that of other reports for optimal growth of bacteria were observed. To some extent, data varied from literature to literature. All these variations may be due to strain variation, host differences, environmental factors, geographical distribution, setting of experiments and extensive sub-culture. The present findings would be helpful for future study to increase the knowledge in bacterial fish pathogens.

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